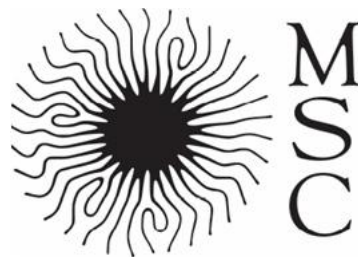


# Effects of brake wear nanoparticles on respiratory cells

Chloé PUISNEY, Evdokia OIKONOMOU, Sophie NOWAK, Alexandre CHEVILLOT,  
Jean-François BERRET and Armelle BAEZA-SQUIBAN

5<sup>th</sup> International NanoSafe Conference, Grenoble, 9<sup>th</sup> November 2016



# General considerations (1)

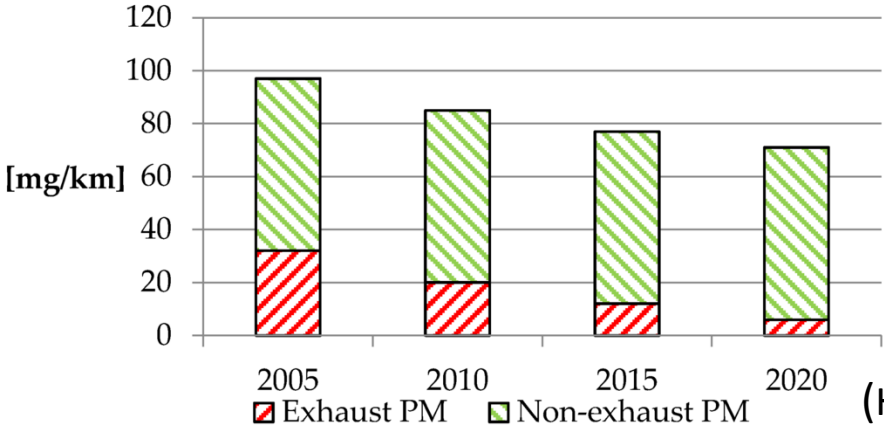
Exhaust



Non-exhaust



Average urban PM10 emissions by passenger cars



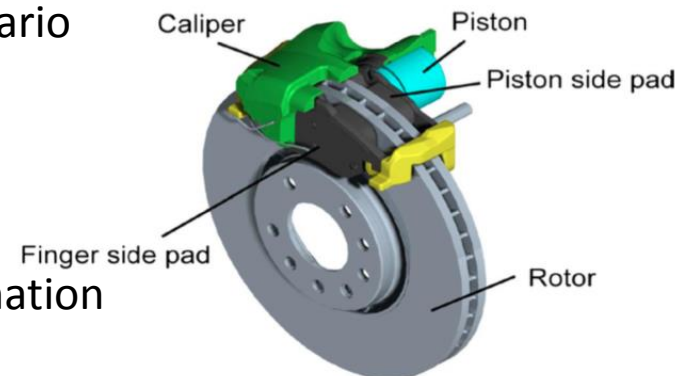
(Hooftman *et al*, Energies, 2016)

- Brake wear contribution (Grigoratos *et al*, Environmental Science Pollution Research, 2015) :
  - up to 55 % by mass to total non-exhaust traffic-related PM<sub>10</sub>
  - up to 21 % by mass to total traffic-related PM<sub>10</sub> emissions

# General considerations (2)

What is already known about brake wear particles ?

- Braking scenario associated with temperature  $\uparrow$  + friction strength
  - Surface chemistry alteration
  - Material degradation
    - Favorable to particles generation
- Wide range of size : from few nanometers to several micrometers  
(Thorpe and Harrisson, Science of Total Environment, 2008)
  - Direct link between particles size and braking scenario
- Due to brake linings composition :
  - Known as non-exhaust linked pollution sources
  - Important source of environmental metal contamination



(Wahlström *et al*, 2009)

# Experimental strategy



# Physicochemical characterization (1)

## X-ray Fluorescence

Chemical element	Composition (%)
Mg	1,6
Al	11,0
Si	13,1
S	6,9
K	2,1
Ca	3,5
Ti	1,3
Cr	1,8
Mn	0,2
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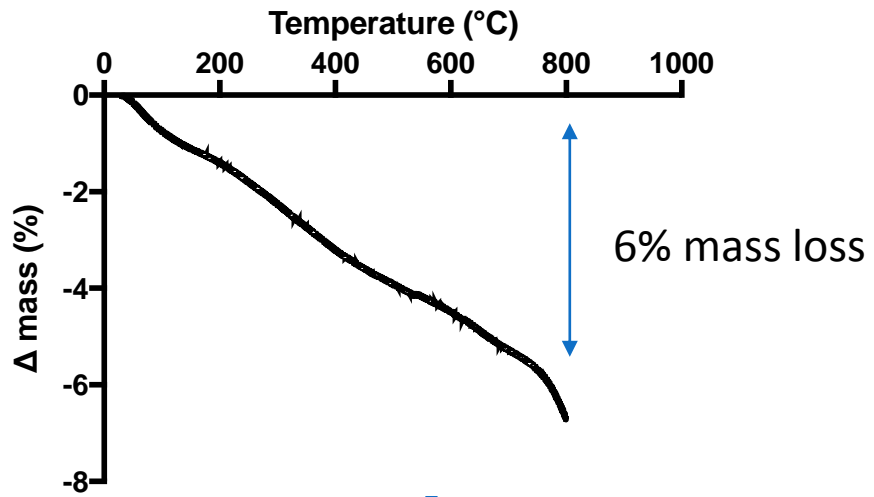
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Thermogravimetric analysis



Organic : 6%  
Inorganic : 94%

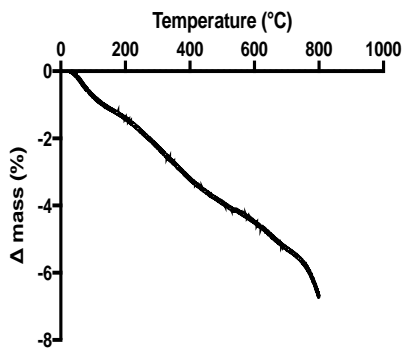


# Physicochemical characterization (1)

X-ray Fluorescence

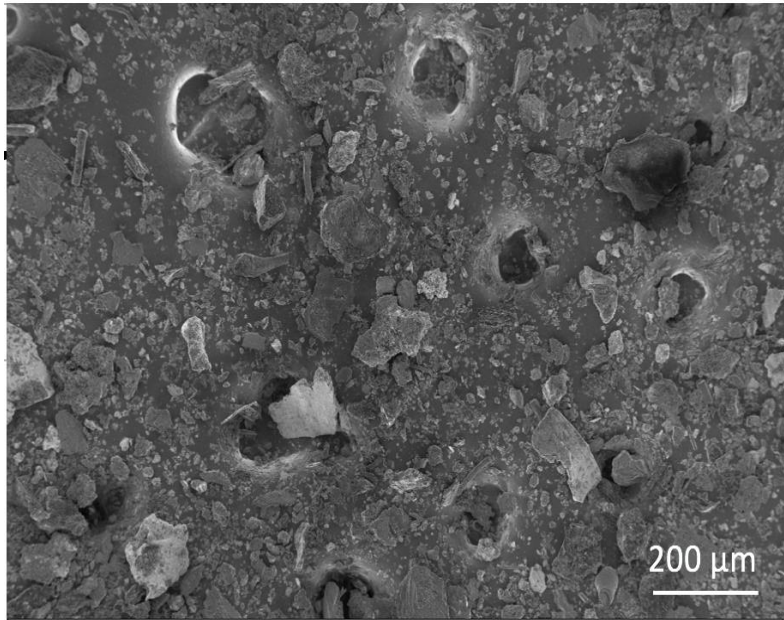
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Scanning Electron Microscopy





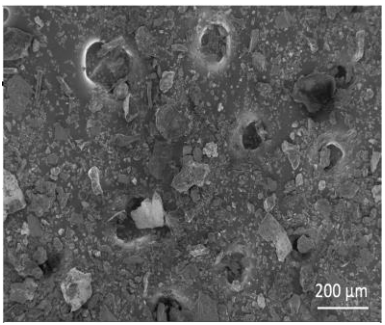
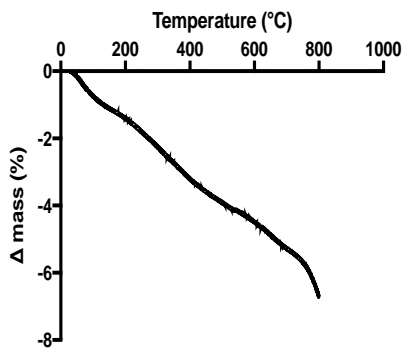
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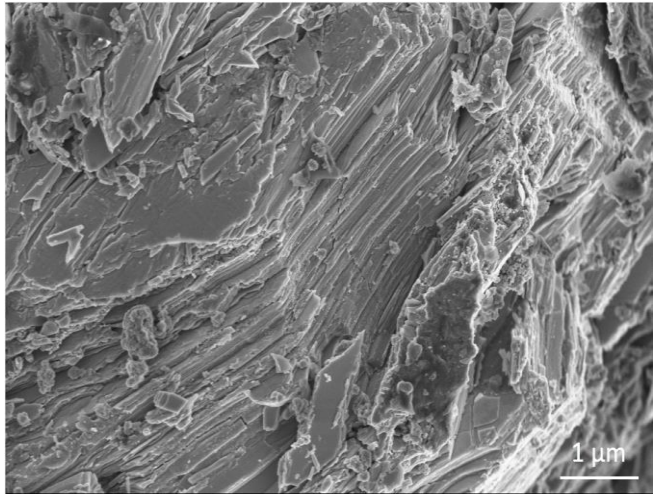
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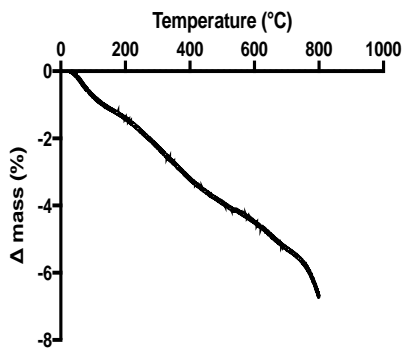


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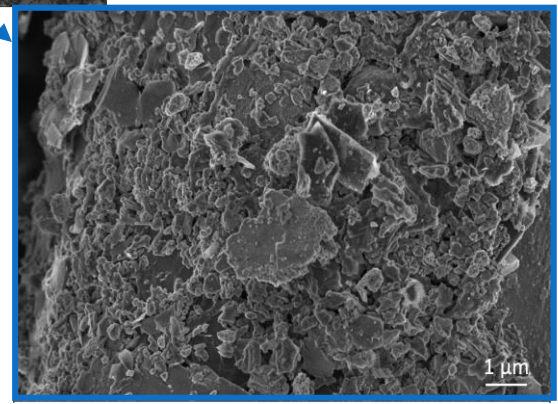
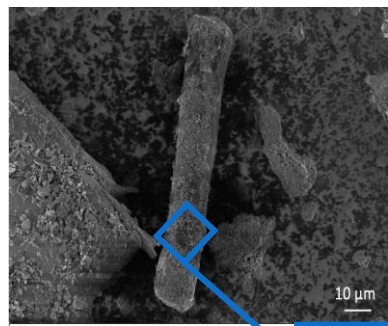
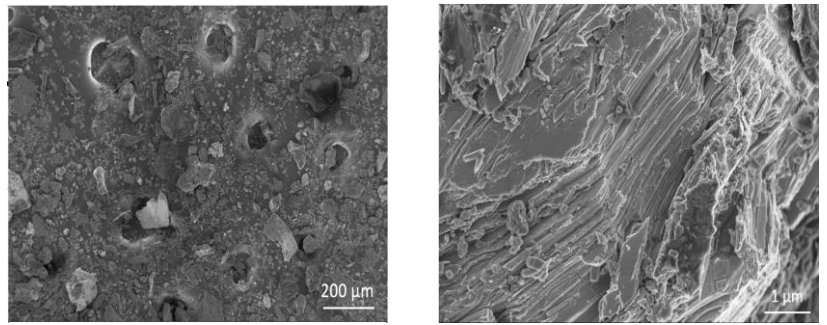
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↓

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## Scanning Electron Microscopy



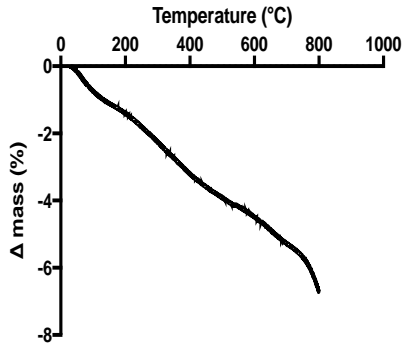
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## X-ray Fluorescence

## Thermogravimetric analysis

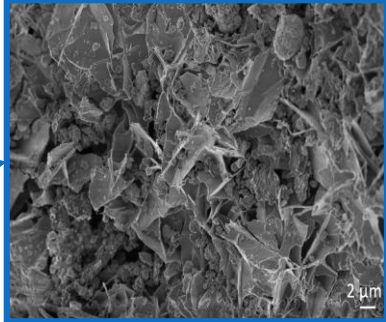
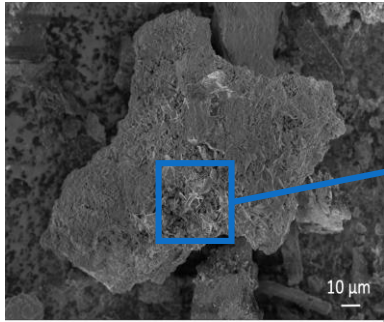
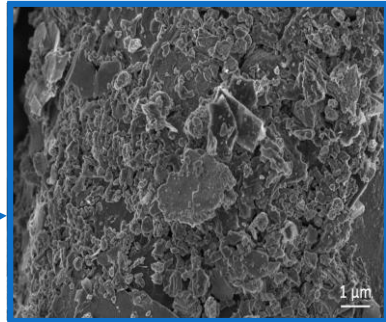
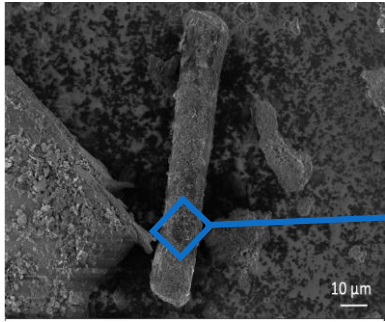
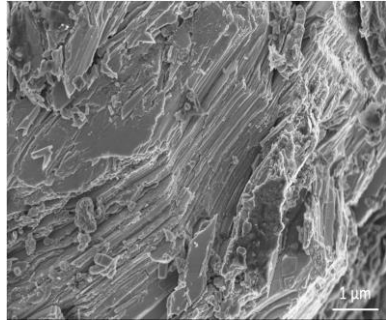
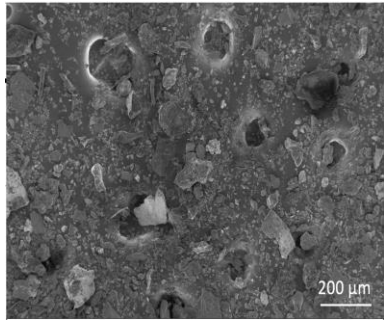
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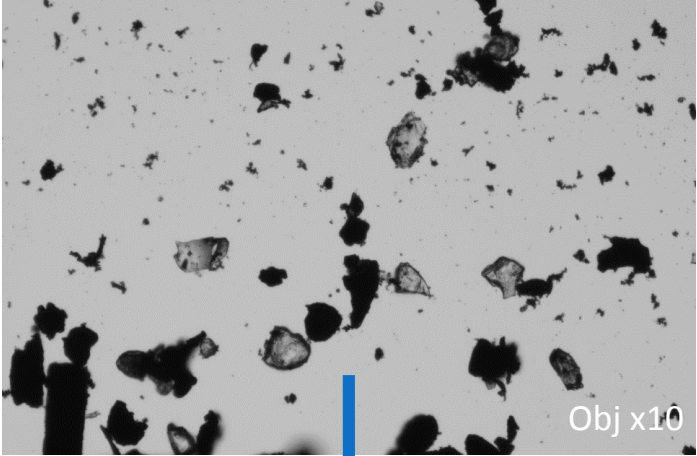
↓

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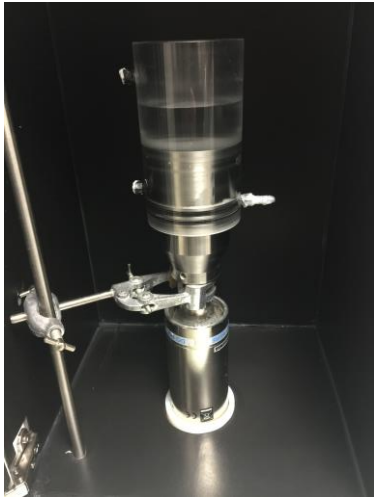


# Physicochemical characterization (2)

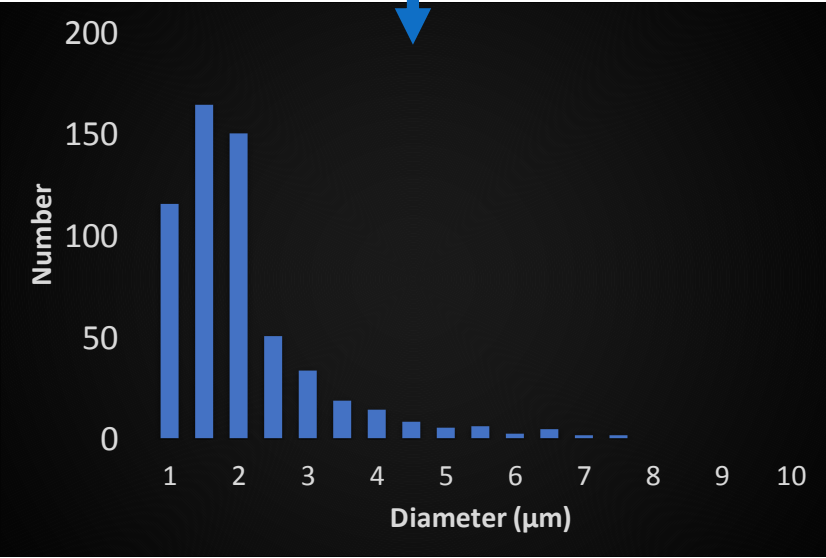
Optical microscopy



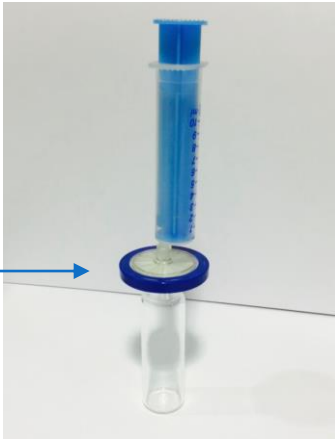
0 → 10 min



Sonication  
+  
Filtration  
process



0,45 µm filter

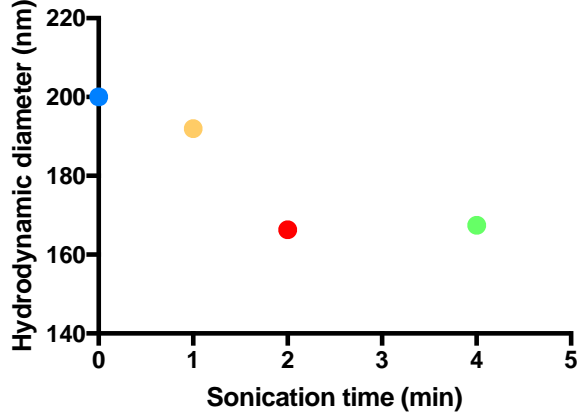
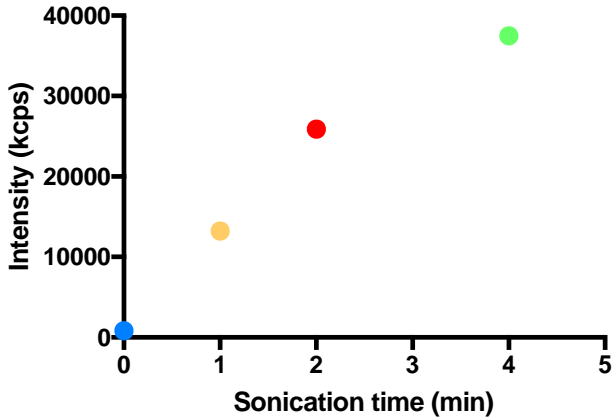




# Physicochemical characterization (3)

## Characterization of brake wear powder suspensions

	Sonication time (min)	Unfiltered	Filtered
Z-average (d.nm)	0	12000	200
	1	1600	192
	2	1200	166
	4	621	167
Intensity (kcps)	0	$3,4 \times 10^5$	$8,1 \times 10^2$
	1	$6,7 \times 10^5$	$1,3 \times 10^5$
	2	$5,2 \times 10^5$	$2,5 \times 10^5$
	4	$1,8 \times 10^6$	$3,7 \times 10^5$



Sonication + filtration process :  
 Size decrease  
 Intensity increase

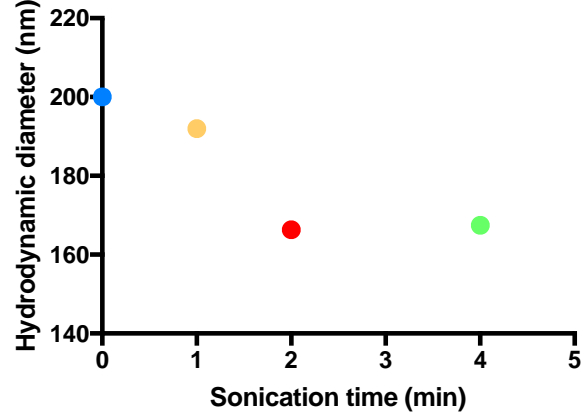
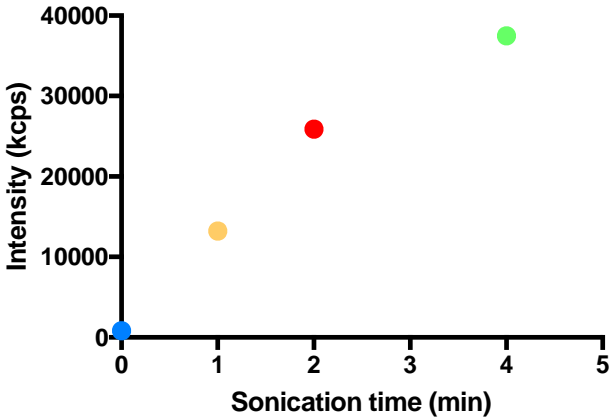


- ✓ Selection of nanometer size fraction
- ✓ Stability study

# Physicochemical characterization (3)

## Characterization of brake wear powder suspensions

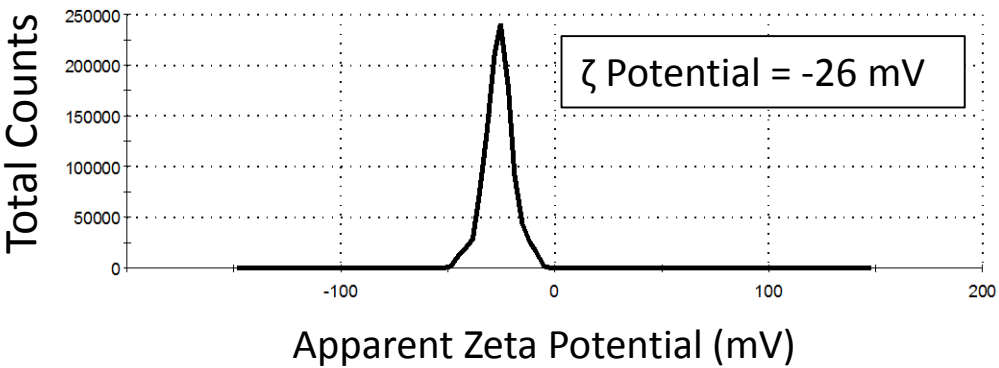
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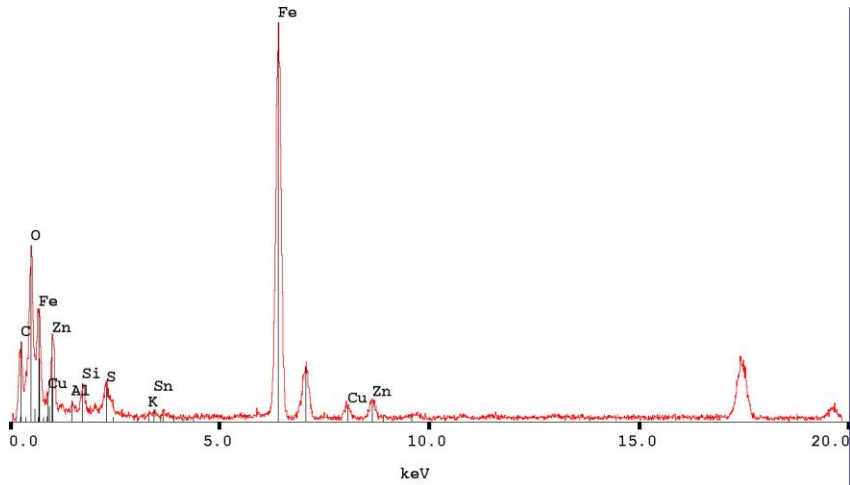
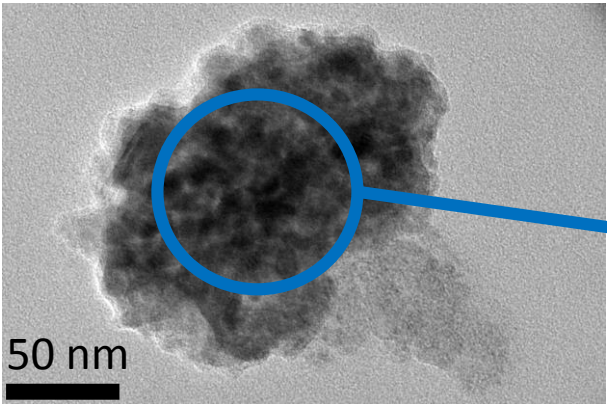
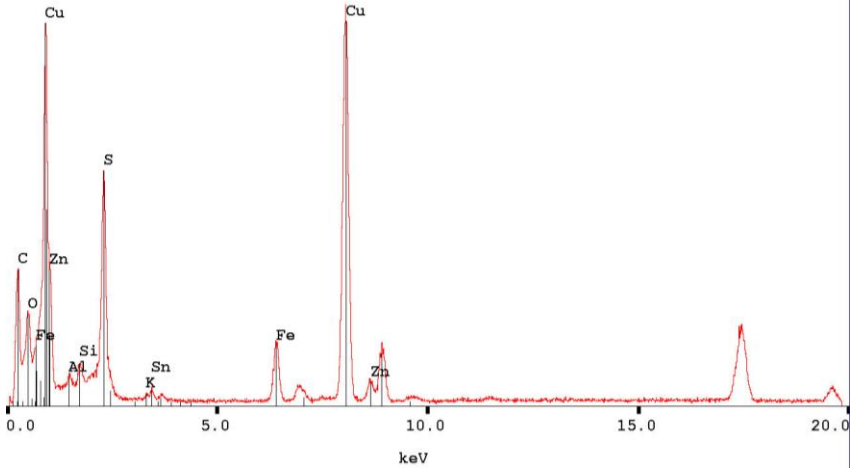
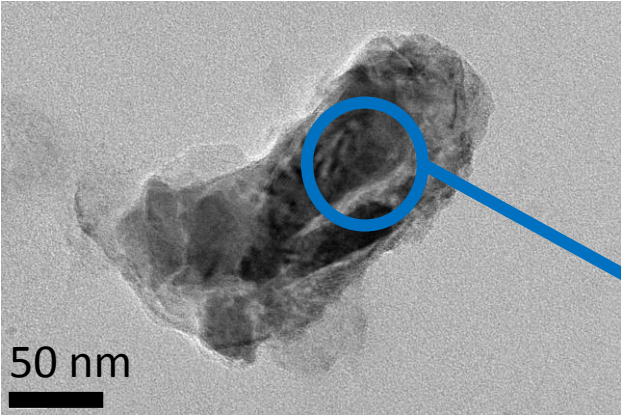


- ✓ Selection of nanometer size fraction
- ✓ Stability study



# Physicochemical characterization (4)

Transmission electron microscopy coupled with energy dispersive X-ray spectroscopy of filtered suspensions



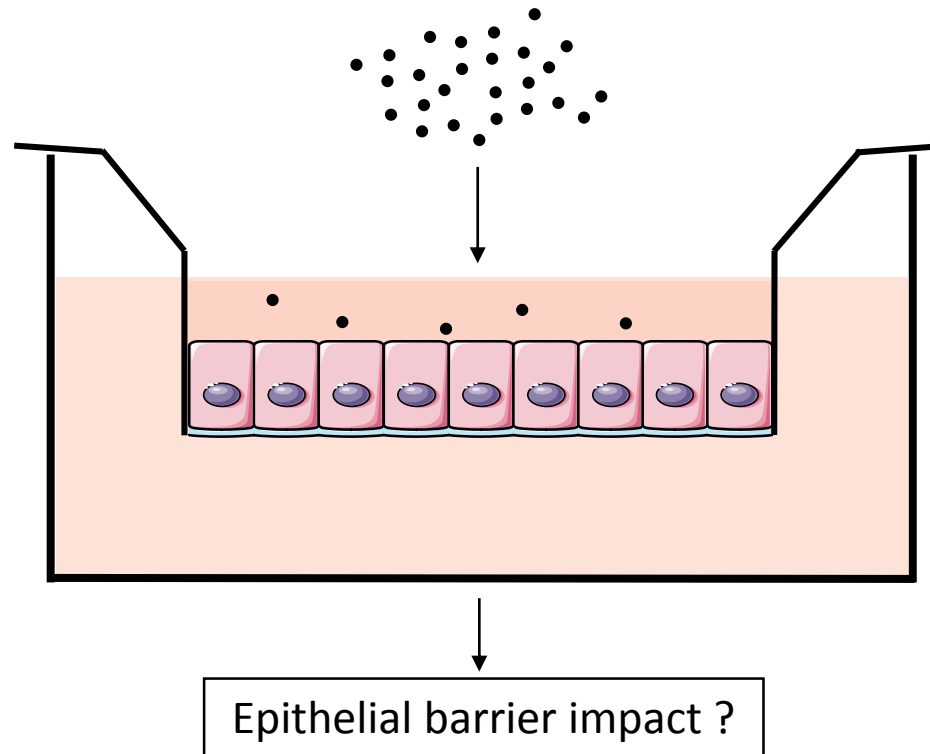


# Toxicological study (1)

24h exposure to :

Brake wear powder suspensions

Benchmark =  $\text{Fe}_2\text{O}_3$  nanoparticles (50 nm)



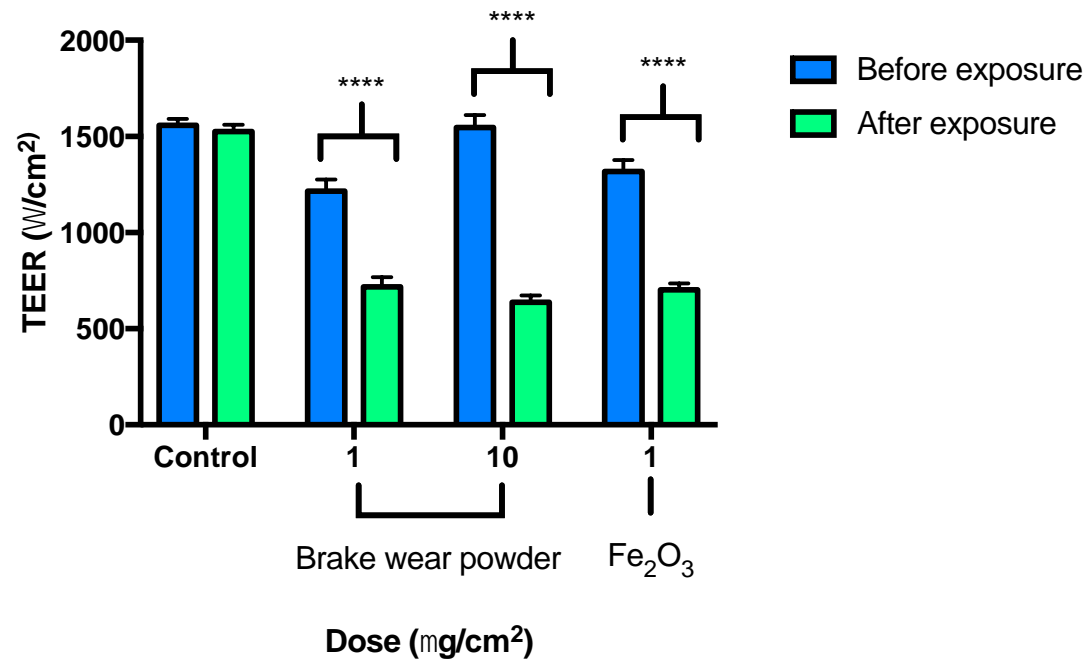
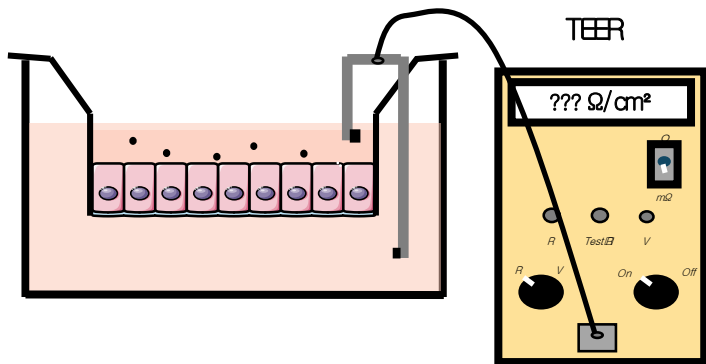
*Calu-3 cells*  
*Human bronchial epithelial cells*  
*TW : Polarized differentiated*  
*epithelium*

- **Barrier Integrity study** : Lucifer Yellow permeability assay, Transepithelial electric resistance measurement (TEER)
- **Cellular morphology** : Immunostaining (Actin, ZO-1, MUC5AC)
- **Proinflammatory response** : ELISA (IL-6, IL-8, TNF)

# Toxicological study (2)

Epithelial barrier integrity assessment following 24h exposure to particles

TEER measurement

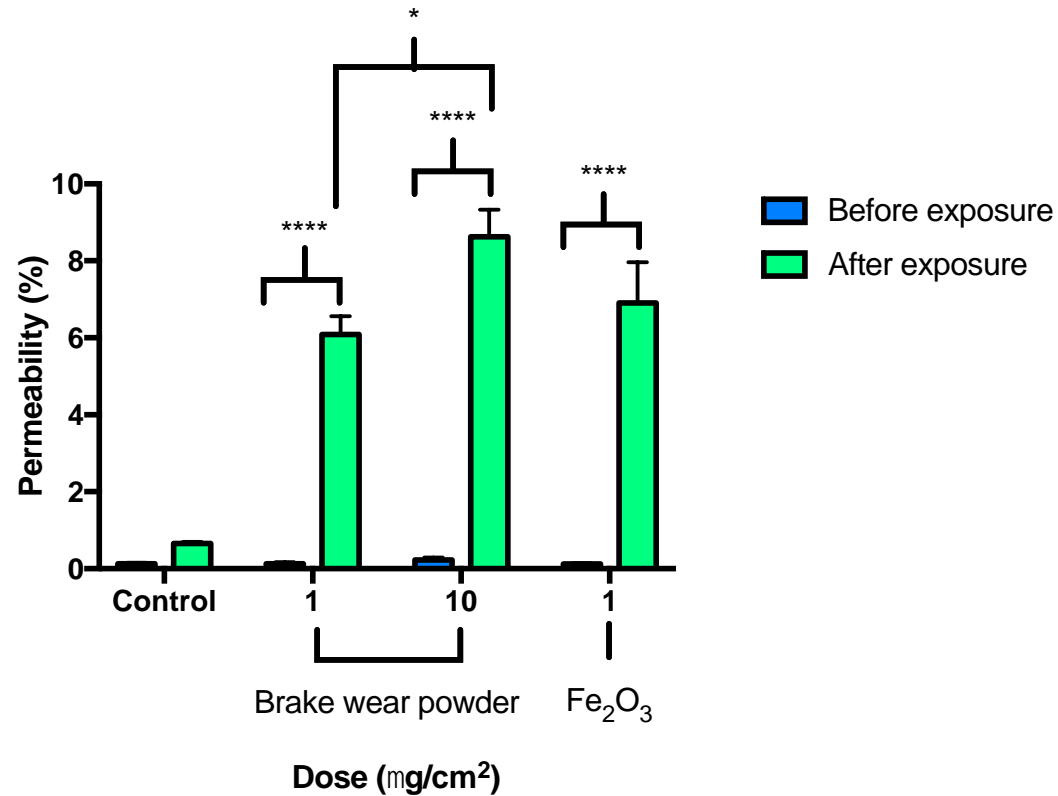
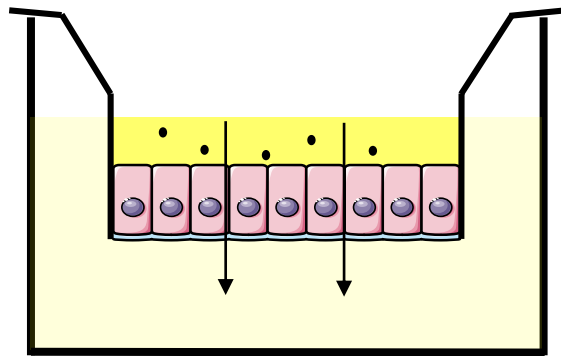


Statistical analysis : ANOVA – Dunnett’s post test, compared to vehicle,  $P < 0,05$  : \*,  $P < 0,01$  : \*\*,  $P < 0,001$  : \*\*\*,  $P < 0,0001$  : \*\*\*\*

# Toxicological study (2)

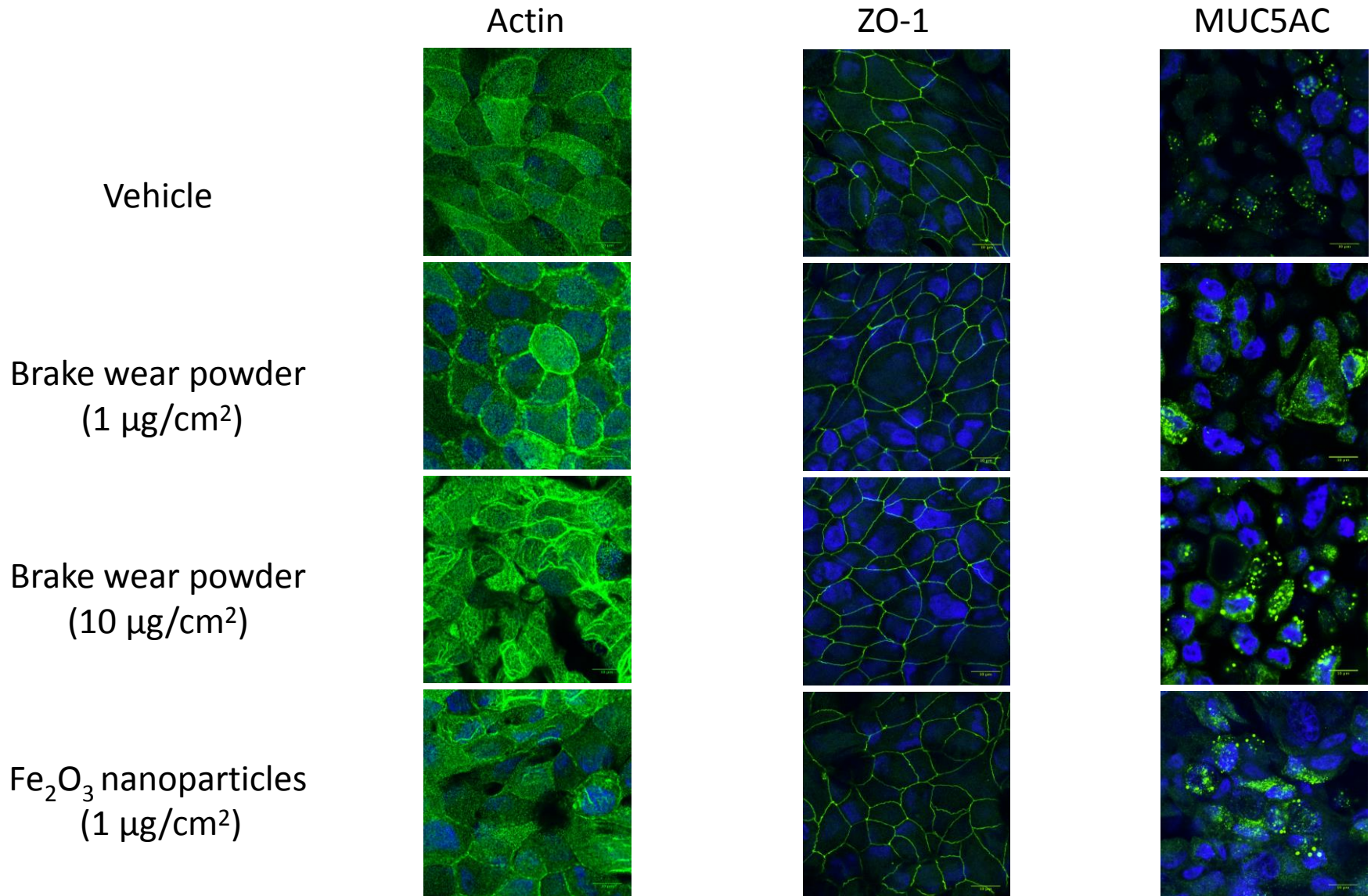
Epithelial barrier integrity assessment following 24h exposure to particles

Lucifer Yellow permeability assay

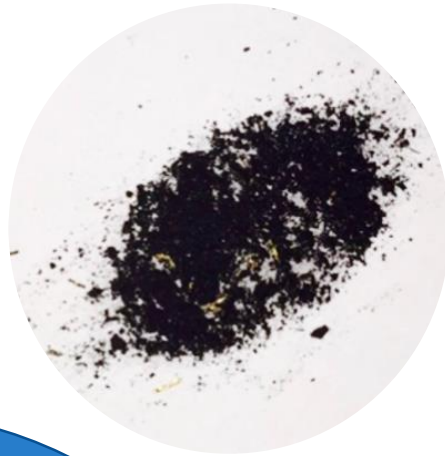


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# Toxicological study (3)



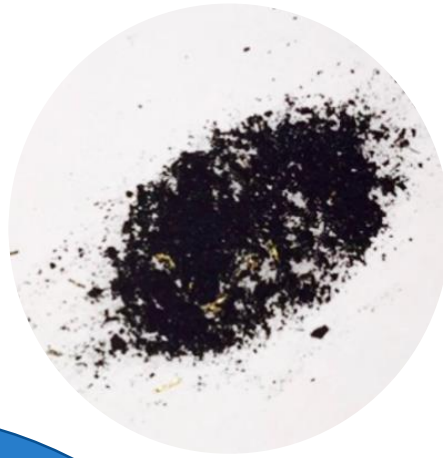
# Conclusions



## Physico-chemical characterization

- Complex mixture
- Major components : Fe, Cu
- Micro- and Nano-sized fractions
- Different amorphous particles

# Conclusions



## Physico-chemical characterization

- Complex mixture
- Major components : Fe, Cu
- Micro- and Nano-sized fractions
- Different amorphous particles

## Toxicological study

- Permeability  $\uparrow$  = Barrier functions  $\searrow$   
= tight junctions injury
- No pro-inflammatory response (data not shown)
- Mucus production  $\uparrow$
- Response similar as benchmark particles

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Fanny MOUSSEAU

Alexandra Sasha LANIECE

Victor BALDIM

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DIM Nano-K, Région Ile-de-France



**UMR CNRS 7086**

**Laboratoire ITODYS**

Alexandre CHEVILLOT

Sophie NOWAK

**Centre d'essais de Juvincourt**

Daniel GOMES

Frédéric RICHARD

**Services de microscopie électronique**

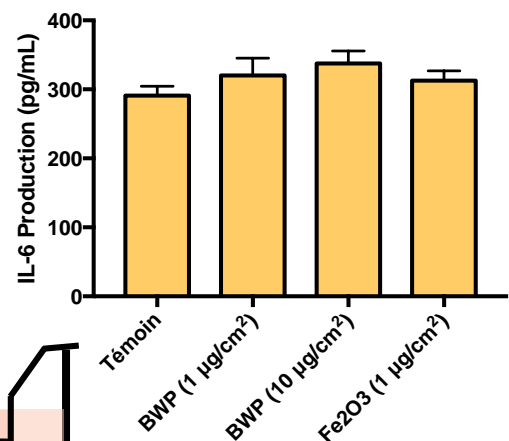
Sandra Casale (UPMC – TEM-EDX)

Ludovic MOUTON (UPD - SEM)

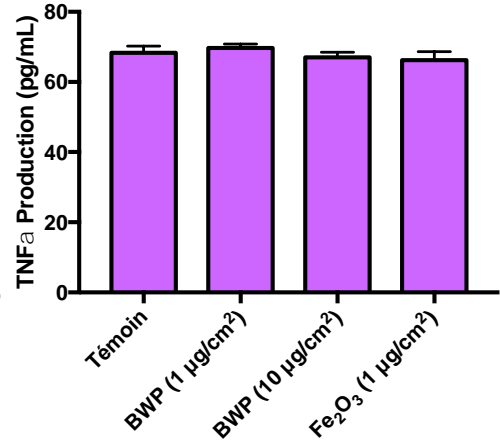


# Pro-inflammatory response

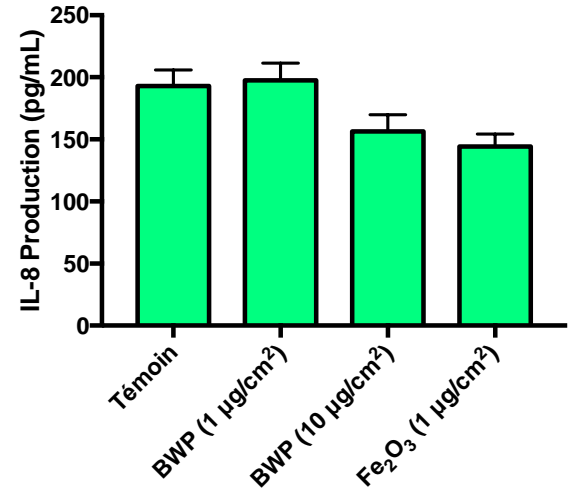
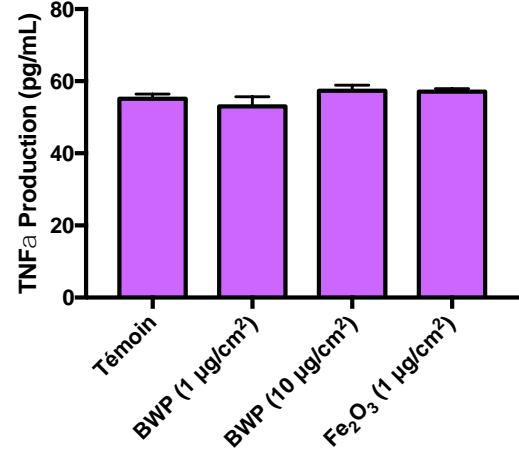
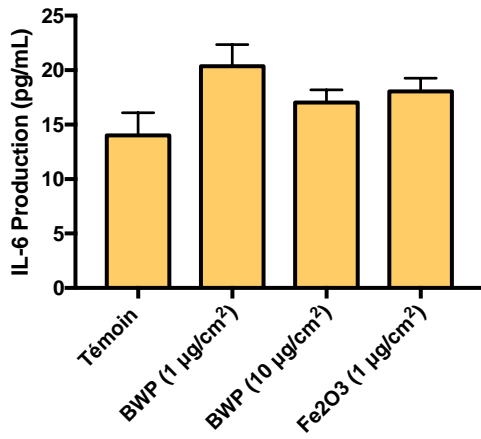
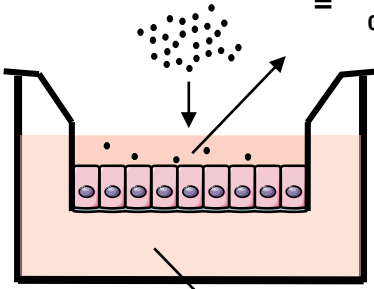
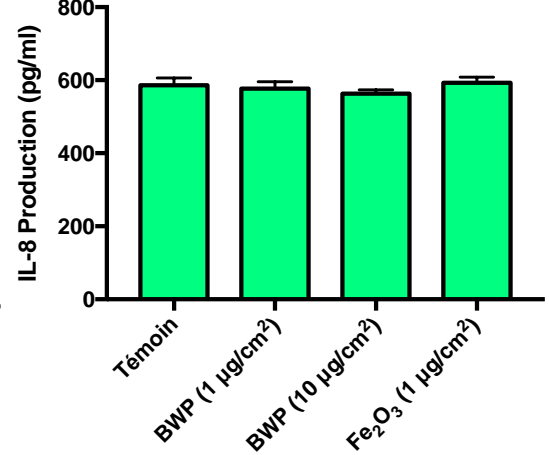
IL-6



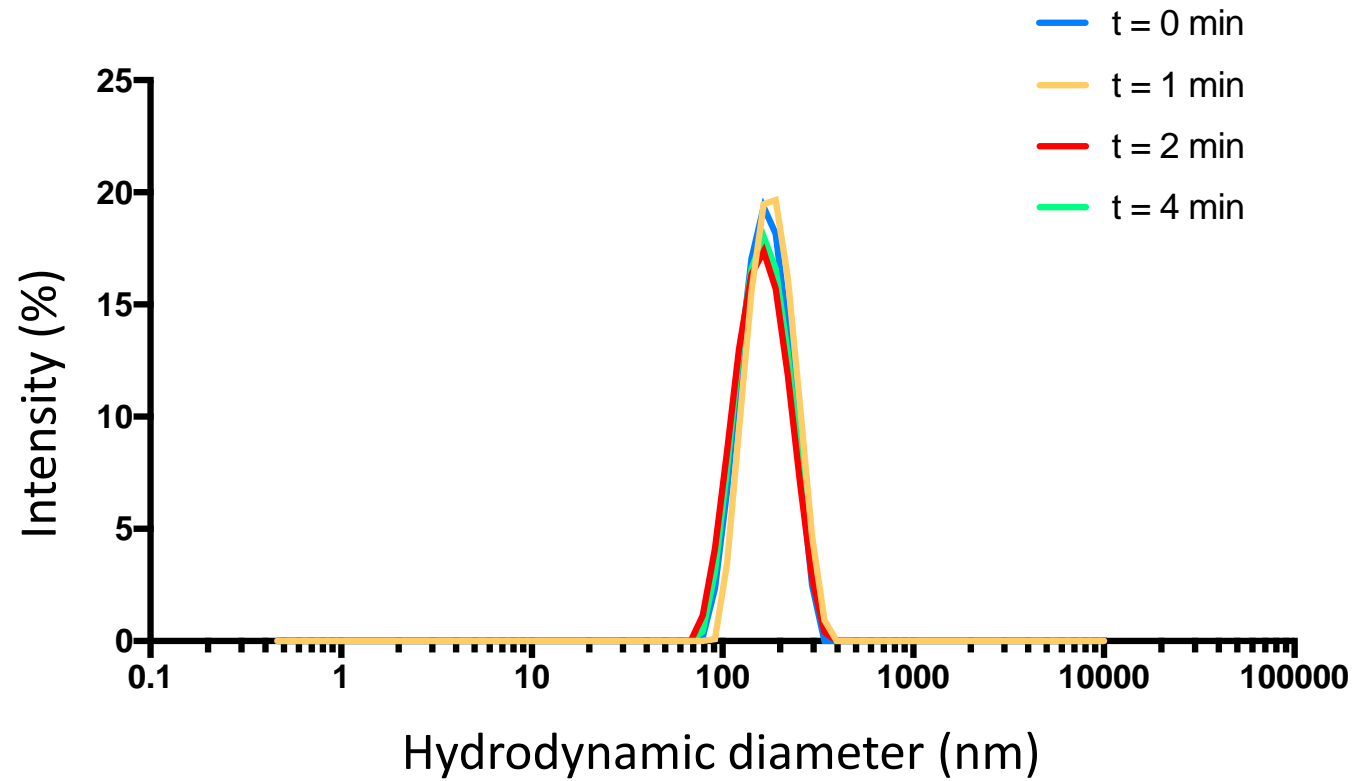
TNF



IL-8



# Size distribution following sonication and filtration process



# Stability study

